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EXAMINER

SAKELARIS, SALLY A

ART UNIT

PAPER NUMBER

1634

DATE MAILED: 04/02/2004

Please find below and/or attached an Office communication concerning this application or proceeding.

Office Action Summary

Application No.

09/869,554

Applicant(s)

ORLEFORS ET AL.

Examiner

Sally A Sakelaris

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-- The MAILING DATE of this communication appears on the cover sheet with the correspondence address --

Period for Reply

A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) FROM THE MAILING DATE OF THIS COMMUNICATION.

- Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication.
- If the period for reply specified above is less than thirty (30) days, a reply within the statutory minimum of thirty (30) days will be considered timely.
- If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication.
- Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133). Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).

Status

- 1) ☒ Responsive to communication(s) filed on 1/20/2004.
- 2a) ☒ This action is **FINAL**. 2b) ☐ This action is non-final.
- 3) ☐ Since this application is in condition for allowance except for formal matters, prosecution as to the merits is closed in accordance with the practice under *Ex parte Quayle*, 1935 C.D. 11, 453 O.G. 213.

Disposition of Claims

- 4) ☒ Claim(s) 2,4,6,12,16 and 19-40 is/are pending in the application.
- 4a) Of the above claim(s) _____ is/are withdrawn from consideration.
- 5) ☐ Claim(s) _____ is/are allowed.
- 6) ☐ Claim(s) 2,4,6,12,16,19-31, and 32-40 is/are rejected.
- 7) ☐ Claim(s) _____ is/are objected to.
- 8) ☐ Claim(s) _____ are subject to restriction and/or election requirement.

Application Papers

- 9) ☐ The specification is objected to by the Examiner.
- 10) ☐ The drawing(s) filed on _____ is/are: a) ☐ accepted or b) ☐ objected to by the Examiner.
Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).
Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.121(d).
- 11) ☐ The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152.

Priority under 35 U.S.C. § 119

- 12) ☐ Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).
- a) ☐ All b) ☐ Some * c) ☐ None of:
- ☐ Certified copies of the priority documents have been received.
 - ☐ Certified copies of the priority documents have been received in Application No. _____.
 - ☐ Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)).

* See the attached detailed Office action for a list of the certified copies not received.

Attachment(s)

- | | |
|--|---|
| 1) <input type="checkbox"/> Notice of References Cited (PTO-892) | 4) <input type="checkbox"/> Interview Summary (PTO-413)
Paper No(s)/Mail Date. _____ |
| 2) <input type="checkbox"/> Notice of Draftsperson's Patent Drawing Review (PTO-948) | 5) <input type="checkbox"/> Notice of Informal Patent Application (PTO-152) |
| 3) <input type="checkbox"/> Information Disclosure Statement(s) (PTO-1449 or PTO/SB/08)
Paper No(s)/Mail Date _____ | 6) <input type="checkbox"/> Other: _____ |

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DETAILED ACTION

This action is in response to Applicant's amendment and response to office action, filed January 20, 2004. Claims 2, 4, 6, 12, 16 and 19 have been amended, claims 1, 3, 5, 7-11, 13-15, and 17-18, have been canceled, and claims 32-40 have been added. Claims 2, 4, 6, 12, 16, and 19-40 are now pending. Applicant's amendments and arguments have been thoroughly reviewed, but are not persuasive for the reasons that follow. All rejections not reiterated herein are hereby withdrawn. The text of those sections of Title 35, U.S. Code not included in this action can be found in a prior Office action. **This action is Final.**

Claim Rejections - 35 USC § 102

The following is a quotation of the appropriate paragraphs of 35 U.S.C. 102 that form the basis for the rejections under this section made in this Office action:

A person shall be entitled to a patent unless –

(b) the invention was patented or described in a printed publication in this or a foreign country or in public use or on sale in this country, more than one year prior to the date of application for patent in the United States.

1. Claims 2, 20 and 21 and new claims 32, 35, and 36 are rejected under 35 U.S.C. 102(b) as being unpatentable over Ronaghi et al. (Anal. Biochemistry, 1996).

Interpreting claim 2's recitation of a "microfluidic device" to mean any device which is suitable to operate with liquids on a microliter scale, Ronaghi et al. teaches the methods of such a device (for example the capillary embodiment on page 88 bottom right).

With respect to claim 2, Ronaghi et al. teach a method of identifying the sequence of a portion of sample DNA comprising the steps of:

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(i) forming immobilized DNA comprising of one strand of sample DNA and one strand of primer DNA on one or more reaction areas in a microchannel structure of a microfluidic device(Pg. 85, bottom right). Incubating the nucleic acid sample with about 0.8 pmol primer, DNA polymerase, and a deoxynucleotide triphosphate(Page 88, Fig. 5).

(ii) adding reagents including deoxynucleotide analogue or dideoxynucleotide and DNA polymerase and moving said reagents within said microchannel structure to each of said one or more reaction areas so that extension of primer occurs as a result from complementarity of the added deoxynucleotide, deoxynucleotide analogue or dideoxynucleotide with the strand of sample DNA that is part of the immobilized double stranded DNA(Page 85- 86)

(iii) detecting whether or not the deoxynucleotide, deoxynucleotide analogue or dideoxynucleotide added in step (ii) is added to the primer DNA in said one or more reaction areas;(Page 86).

(iv) removing reagents such as deoxynucleotide from one or more reaction areas; is taught throughout the Ronaghi reference in their teachings in Figure 1 and later on page 87 as they wash the beads on which the deoxynucleotides are immobilized, the reference further teaches the loss of these deoxynucleotides following the wash steps on page 87.

(v) repeating steps (ii)-(iv) with different deoxynucleotides, deoxynucleotide analogues or dideoxynucleotides is taught by Ronaghi in Figure 1 and in the text of Page 87 in their teaching that “the sequencing procedures were repeated several times”.

(vi) identifying said sequence from the results of the above previous steps is obviously then taught in the reference’s sequencing previously alluded to in (v) and furthermore that “the

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obtained sequence was confirmed by semiautomated solid-phase Sanger sequencing”(Pg. 87, see figure 5).

With regard to claim 20, Ronaghi et al. teaches the above method wherein the detecting step (iii) measures the release of pyrophosphate(Page 85).

With regard to claim 21, Ronaghi et al further teach the method wherein the pyrophosphate release is detected by light emitted from a luciferin luciferase reaction(Fig. 1, Pg. 85).

With regard to new claim 32, Ronaghi et al. further teaches the method of claim 2 wherein step (iv) is washing one or more reaction areas in their teaching on the left side of page 85 of “washing of the immobilized DNA fragments between each nucleotide addition was performed”(Ronaghi pg. 85). Additionally the reference teaches in the abstract that the “parallel processing of many samples in an automated manner is discussed”.

With regard to claims 35 and 36, the reference anticipates the limitations of the method of claim 2 and claim 35 wherein the amount of DNA sample is in the range of about 1 femtomole to about 200 pmole and also about 0.1 pmol to about 200pmol in their teaching on page 85 of “one picomole of the immobilized DNA fragment” being used in the sequencing reaction.

Response to Arguments:

Applicant's arguments filed 1/20/2004 have been fully considered but they are not persuasive. While applicant's arguments that it is well known to those of skill in the art that the term “microfluidics” and “microfluidic device” refer to a device in which there is a transport of liquid, and their reference to page 5, line 32 to page 6, line 8 and Figure 1 is acknowledged, these arguments are not found to be convincing. The applicant asserts that “microfluidic devices

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require transport of sample, reagents and the like by the use of liquid flow”(response pg. 9).

Applicant is pointed to page 88 of the Ronaghi reference and their teaching of their immobilization of the DNA template in a capillary and their further characterization of such a system as “a flow system, with small volumes, high speed”, which meets applicant’s own standard of a characteristic of a microfluidic device. In addition applicant argues that their new limitation to indicate that fluids are moved within the microchannel structure is not taught by Ronaghi et al. Although this amendment’s new limitation is acknowledged, it does not make the claim free of the cited prior art. Again, applicant is pointed to the “flow system”(pg. 88) that is described in the Ronaghi reference that anticipates this new limitation of moving in the microchannel structure.

Claim Rejections - 35 USC § 103

The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negated by the manner in which the invention was made.

2. Claims 2, 4, 6, 12, 16, 19-31 and new claims 33, 34, and 37-40 are rejected under 35 U.S.C. 103(a) as being unpatentable over Ronaghi et al.(Anal. Biochemistry, 1996) in view of Mian et al.(US Patent 6,319,469 B1).

Interpreting claim 2’s recitation of a “microfluidic device” to mean any device which is suitable to operate with liquids on a microliter scale, Ronaghi et al. teaches the methods of such a device(for example the capillary embodiment on page 88 bottom right).

With respect to claim 2, Ronaghi et al. teach a method of identifying the sequence of a portion of sample DNA comprising the steps of:

- (i) forming immobilized DNA comprising of one strand of sample DNA and one strand of primer DNA on one or more reaction areas in a microchannel structure of a microfluidic device(Pg. 85, bottom right). Incubating the nucleic acid sample with about 0.8 pmol primer, DNA polymerase, and a deoxynucleotide triphosphate(Page 88, Fig. 5).
- (ii) adding reagents including deoxynucleotide analogue or dideoxynucleotide and DNA polymerase and moving said reagents within said microchannel structure to each of said one or more reaction areas so that extension of primer occurs as a result from complementarity of the added deoxynucleotide, deoxynucleotide analogue or dideoxynucleotide with the strand of sample DNA that is part of the immobilized double stranded DNA(Page 85- 86)
- (iii) detecting whether or not the deoxynucleotide, deoxynucleotide analogue or dideoxynucleotide added in step (ii) is added to the primer DNA in said one or more reaction areas;(Page 86).
- (iv) removing reagents such as deoxynucleotide from one or more reaction areas; is taught throughout the Ronaghi reference in their teachings in Figure 1 and later on page 87 as they wash the beads on which the deoxynucleotides are immobilized, the reference further teaches the loss of these deoxynucleotides following the wash steps on page 87.
- (v) repeating steps (ii)-(iv) with different deoxynucleotides, deoxynucleotide analogues or dideoxynucleotides is taught by Ronaghi in Figure 1 and in the text of Page 87 in their teaching that “the sequencing procedures were repeated several times”.

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(vi) identifying said sequence from the results of the above previous steps is obviously then taught in the reference's sequencing previously alluded to in (v) and furthermore that "the obtained sequence was confirmed by semiautomated solid-phase Sanger sequencing"(Pg. 87).

With regard to claim 20, Ronaghi et al. teaches the above method wherein the detecting step (iii) measures the release of pyrophosphate(Page 85).

With regard to claim 21, Ronaghi et al further teach the method wherein the pyrophosphate release is detected by light emitted from a luciferin luciferase reaction(Fig. 1, Pg. 85).

With regard to new claim 32, 33, and 34 Ronaghi et al. further teaches the method of claim 2 wherein step (iv), claim 4 wherein step (vi), and claim 19, wherein step (vii) is washing one or more reaction areas in their teaching on the left side of page 85 of "washing of the immobilized DNA fragments between each nucleotide addition was performed"(Ronaghi pg. 85). Additionally the reference teaches in the abstract that the "parallel processing of many samples in an automated manner is discussed".

With regard to claims 35-40 the reference anticipates the limitations of the method of claim 2, 35, 4, 19,37, and 39 wherein the amount of DNA sample is in the range of about 1 femtomole to about 200 pmole and also about 0.1 pmol to about 200pmol in their teaching on page 85 of "one picomole of the immobilized DNA fragment" being used in the sequencing reaction.

But, with respect to Claims 4, 6, 12, 16, 19, and 22-31 Ronaghi et al. does not teach a method for identifying the sequence of a portion of sample DNA wherein the steps are performed in a microfluidic device that is a disk and the fluids are moved(claims 4, 12, 16, and

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19) by centripetal force, such as that which is referred to on page 5, line 32 of the current specification. Furthermore, Ronaghi et al. does not teach labeling the deoxynucleotide, deoxynucleotide analogue, or dideoxynucleotide that is added in the method.

However, Mian et al. (US Patent 6,319,469 B1) teach performing the previously taught methods of Ronaghi inside another type of microfluidic device. Mian et al. teach performing the steps of adding sample DNA on a reaction area in a microfluidic device(see Col. 49 lines 1-4), attaching or hybridizing single stranded DNA, and plainly adding sample DNA to a predetermined area on a microfluidic device that is a disc and whose fluids can be moved to various chambers(Col. 49 lines 2-19). Furthermore, the Mian et al. reference adds teachings of a disc-shaped, microfluidic device that causes fluid movement through the use of centripetal force(Col. 3 lines 5-25). The reference even further teaches that such methods and apparatus are advantageous as they fill the need in the art for a “simple, flexible, reliable, rapid, and economical microanalytic and microsynthetic reaction platform for performing biological, biochemical, and chemical analyses and syntheses that can move nanoliter to microliter amounts of fluids”(Col. 3 lines 5-10). The reference provides that the invention also advantageously combines “wet” chemistry capabilities with information processing, storing and manipulating ability. The addition of the disc-shaped microfluidic device that exploits centripetal force, to this method for sequence identification, conferred the ability to properly mix reaction components, remove reaction side products, and isolate desired reaction products and intermediates.(Col 3, lines 5-25)(Col 48, line 67) Furthermore, Mian et al. add the teaching of forming DNA to a “microchannel structure” within the microfluidic device. The reference teaches that; the unique disc shape and ability to move nanoliter to microliter amounts of fluid, including reagents and

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reactants, at rapid rates to effect the proper mixing of reaction components through the use of microchannel structures and centripetal force, provides a remedy for the many deficiencies of the status quo. The use of microchannels, functioning to separate micro-amounts of fluid reagents, and centripetal force, to move fluids into and out of reaction chambers, facilitates high-throughput analysis for both genome sequencing and routine clinical applications “that are sophisticated(for professional, eg hospital, use), easy to use(for consumer eg at-home monitoring, uses), and portable (for field environmental testing, use)” (Col. 3 lines 19-22).

Therefore, it would have been obvious to one of ordinary skill in the art at the time the invention was made to have conducted the method of Ronaghi et al. in view of the methods of Mian et al. by incorporating a disc-shaped microfluidic device with microchannels and caused fluid flow through the use of centripetal force in order to have achieved the expected benefit of providing a method that could be used for the automation of larger sequencing projects and for the provision of a “high-throughput system.”

With respect to Claims 6, 22, 25, 26, and 29-31 and the limitation of a fluorescently labeled dideoxynucleotide, Mian teaches a detection step that involves a labeled terminator (Col 49, lines 5-10). Mian et al. teach a method wherein the detection step comprises the DNA being transferred into a mixing chamber containing terminator solution by spinning the disk(Col. 47 lines 15, 28, 39 for example). Terminator solution typically comprises 100nl of a solution containing 5 picomoles of each deoxynucleotide and 0.5 picomoles of one dideoxynucleotide covalently linked to a fluorescent label. The set of dideoxynucleotide-terminated DNA fragments comprising the reaction mixture is then separated by capillary electrophoresis and the sequence of the fragments determined by laser-induced fluorescence detection. The reference

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further teaches that this mode of detection ie, discs comprising a multiplicity of these synthetic arrays with fluorescent labels, permits the simultaneous synthesis of a plurality of dideoxynucleotide-terminated oligonucleotides and therefore applicable in high throughput analysis of sequencing data or clinical approaches. Mian et al. teaches the use of a terminator solution containing a dideoxynucleotide covalently-linked to a fluorescent label in Example 7, Col. 49. In addition, Mian et al. teach, in addition to the aforementioned, fluorescently labeled dideoxynucleotide of Example 7, Example 3 which includes the incorporation of fluorescently labeled DNA to one or more reaction areas so that extension of primer occurs as a result from complementarity of the added dideoxynucleotides with the strand of sample DNA that is part of the immobilized double stranded DNA.

Therefore, it would have been obvious to one of ordinary skill in the art at the time the invention was made to have conducted the method of Ronaghi et al. in view of the methods of Mian et al. and to have added a labeled terminator and fluorescently labeled dideoxynucleotides, in order to have achieved the benefit of providing a method that, would permit the simultaneous synthesis of a plurality of fluorescently labeled dideoxynucleotide-terminated oligonucleotides and therefore applicable in high throughput analysis of sequencing data or clinical approaches.

Response to Arguments:

Applicant's arguments filed 1/20/2004 have been fully considered but they are not persuasive. While applicant's arguments "that neither Ronaghi et al. not Mian et al. alone or in combination teaches and/or suggests the present invention" are acknowledged they are not found to be convincing. First applicant asserts that Mian et al. teaches in Example 7 (col.48-49) a different sequencing procedure from that taught by Ronaghi et al. However, applicant is reminded that

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both methods are sequencing methods that contain deoxynucleotides and a DNA polymerase, which are the only two reagents that are required by the present claims as being necessary for carrying out a sequencing reaction. As a sequencing method is presently being defined by the claims, both references teach the same sequencing method and are appropriately combined in the obviousness rejection.

Applicant next asserts that the two references teach a different method since the Mian et al. reference does not teach “immobilization of DNA for sequencing”. Applicant is pointed to Col. 42 lines 46-48 where the Mian et al. reference teaches the immobilization of one strand of DNA on the disk by attaching it to a streptavidin coated bead. Which, is the very method taught to be used by applicants in their specification on page 7. As such the reference is not seen as teaching away from the invention, but contrastingly that it specifically suggests the applicant’s method as presently claimed and as a result has been applied appropriately in the presently maintained obviousness rejection. Furthermore while a motivation was provided in the last office action(pgs 8 and 9) additional motivation for immobilizing the DNA as taught in Example 3 of the Mian et al. reference is that “user preparation of the DNA is minimized and the cost of DNA preparation per sample is greatly reduced”(Col. 42 lines 62 and 63).

Next, applicant argues that the “examiner appears to pick and choose elements from the various DNA procedures taught by Mian et al. as a suggestion to combine the Ronaghi et al. and Mian et al.”. Applicants argue that Mian et al. does not teach adding dideoxy nucleotides in a sequencing reaction. However applicant is pointed to example 7 in Mian et al. where in Col. 49, Mian teach a sequencing method using “a solution comprising a dideoxy form of nucleotides G, A, T or C”(lines 4-5). The examiner is not picking and choosing as the sequencing example

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expressly teaches the use of dideoxy nucleotides. In response to applicant's argument that the washing of reagents is taught only in the context of a DNA synthesis method, the rejection's primary reference, Ronaghi et al. teaches washing between each nucleotide addition on page 85. The claims are met by the references cited. One of ordinary skill in the art would understand that many of these limitations (immobilization of DNA, washing or removing reagents, the use of dideoxys) can be used in different methods to achieve the same result, the title of the method does not preclude the use of the same reagents that are equally applicable to various DNA based methods and whose application as such would be obvious to one of ordinary skill in the art at the time the invention was made. Applicant next argues that the previously made rejections were an application of an "obvious to try" standard. Applicant is reminded that:

The legal standard for "reasonable expectation of success" is provided by case law and is summarized in MPEP 2144.08, which notes "obviousness does not require absolute predictability, only a reasonable expectation of success; i.e., a reasonable expectation of obtaining similar properties. See, e.g., *In re O'Farrell*, 853 F.2d 894, 903, 7 USPQ2d 1673, 1681 (Fed. Cir. 1988)." In this factual case, there is express suggestion in the prior art that real time sequencing has been performed in a microfluidic device in Ronaghi et al. (Title and capillary embodiment of pg. 88) teaching of "Real Time DNA Sequencing Using Detection of Pyrophosphate Release" article featuring the methods use in embodiments that immobilize the DNA template in a capillary, which had been previously described as a microfluidic device. There is further evidence as shown in the Mian et al. reference in their teaching of "real time control capabilities are also provided" (Col. 34 line 40). This is sufficient for a reasonable

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expectation of success. The MPEP cites *In re O'Farrell*, which notes regarding "obvious to try" at page 1682, that,

"In some cases, what would have been "obvious to try" would have been to vary all parameters or try each of numerous possible choices until one possibly arrived at a successful result, where the prior art gave either no indication of which parameters were critical or no direction as to which of many possible choices is likely to be successful. E.g., *In re Geiger*, 815 F.2d at 688, 2 USPQ2d at 1278; *Novo Industri A/S v. Travenol Laboratories, Inc.*, 677 F.2d 1202, 1208, 215 USPQ 412, 417 (7th Cir. 1982); *In re Yates*, 663 F.2d 1054, 1057, 211 USPQ 1149, 1151 (CCPA 1981); *In re Antonie*, 559 F.2d at 621, 195 USPQ at 8-9. In others, what was "obvious to try" was to explore a new technology or general approach that seemed to be a promising field of experimentation, where the prior art gave only general guidance as to the particular form of the claimed invention or how to achieve it. *In re Dow Chemical Co.*, 837 F.2d, 469, 473, 5 USPQ2d 1529, 1532 (Fed. Cir. 1985); *Hybritech, Inc. v. Monoclonal Antibodies, Inc.*, 802 F.2d 1367, 1380, 231 USPQ 81, 90-91 (Fed. Cir. 1986), cert. denied, 107 S.Ct. 1606 (1987); *In re Tomlinson*, 363 F.2d 928, 931, 150 USPQ 623, 626 (CCPA 1966).

The court in *O'Farrell* then, affirming the rejection, notes "Neither of these situations applies here." For the instant case, it is clear that neither situation applies here either. This is not a situation where the prior art suggests varying a variety of parameters, since the prior art directly points to real time sequencing being performed in a microfluidic device in both the Ronaghi et al. reference and the Mian et al reference. This is also not a situation where only general guidance was given. The prior art provides specific guidance directing real time sequencing in a microfluidic device.

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New Matter

5. Claims 2, 4, 6, 12, 16, and 19-40 are rejected under 35 U.S.C. 112, first paragraph, as containing subject matter which was not described in the specification in such a way as to reasonably convey to one skilled in the relevant art that the inventor(s), at the time the application was filed, had possession of the claimed invention. MPEP 2163.06 notes "If new matter is added to the claims, the examiner should reject the claims under 35 U.S.C. 112, first paragraph - written description requirement. In re Rasmussen , 650 F.2d 1212, 211 USPQ 323 (CCPA 1981)."

In the instantly rejected claims, the new limitation of "removing said reagents" (said referring to deoxynucleotide, deoxynucleotide analogue, dideoxynucleotide, and DNA polymerase in step (ii)) in claims 2, 4, and 19 appears to represent new matter. No specific basis for this limitation was identified in the specification, nor did a review of the specification by the examiner find any basis for the limitation. Since no basis has been identified, the claims are rejected as incorporating new matter. While basis was found on pages 2 and 8 of the specification for the inclusion of a step that removes or washes excess deoxynucleotide or deoxynucleotide analogue, no basis was found for the other elements, and applicant provided no guidance as to the basis for their amendment.

Response to Arguments:

Applicant's arguments filed 1/20/2004 have been fully considered but they are not persuasive. While examiner acknowledges that the insertion of the phrases cited by applicant from their specification including; "solution is removed from the well"(pg. 2 line 2), page 8 line 33's recitation of a washing step, removal of a "solution" or "liquid", or the recitation on page 9 lines

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1-5 “the washing step” all would **not** be considered as representing new matter, the present recitation of “removing said reagents” is as basis for all it encompasses neither has been found by the examiner nor pointed to by applicant’s representative and as such the rejection is maintained. The problem is not washing of dNTPs, but rather the washing of all of the other “said reagents”. The specification only provides basis for the removal or washing away of the deoxynucleotides on pages 2 and 8, no basis was found for the washing or removal of dideoxynucleotides or DNA polymerase.

Applicant's amendment necessitated the new ground(s) of rejection presented in this Office action. Accordingly, **THIS ACTION IS MADE FINAL**. See MPEP § 706.07(a).

Applicant is reminded of the extension of time policy as set forth in 37 CFR 1.136(a).

A shortened statutory period for reply to this final action is set to expire **THREE MONTHS** from the mailing date of this action. In the event a first reply is filed within **TWO MONTHS** of the mailing date of this final action and the advisory action is not mailed until after the end of the **THREE-MONTH** shortened statutory period, then the shortened statutory period will expire on the date the advisory action is mailed, and any extension fee pursuant to 37 CFR 1.136(a) will be calculated from the mailing date of the advisory action. In no event, however, will the statutory period for reply expire later than **SIX MONTHS** from the date of this final action.

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Any inquiry concerning this communication or earlier communications from the examiner should be directed to Sally A Sakelaris whose telephone number is 571-272-0748. The examiner can normally be reached on M-Fri, 9-6:30.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Gary Benzion can be reached on 571-272-0782. The fax phone number for the organization where this application or proceeding is assigned is 703-872-9306.

Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see <http://pair-direct.uspto.gov>. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free).

Sally Sakelaris

4/1/2004



JEFFREY FREDMAN
PRIMARY EXAMINER